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**Title: FLUORESCENCE POLARIZATION-BASED
HOMOGENEOUS ASSAY FOR DEOXYNIVALENOL
DETERMINATION IN GRAINS**

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Inventors: Mohammad Sarwar Nasir
1012 Highgate Lane
Grayslake, Illinois 60030

Citizen of U.S.A.

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Michael E. Jolley
34469 North Circle Drive
Round Lake, Illinois 60073

25

Citizen of United Kingdom

1012 Highgate Lane
Grayslake, Illinois 60030

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application No. 60/231,887, filed on September 11, 2000.

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BACKGROUND OF THE INVENTION

1. Field of the Invention

This invention relates to the field of mycotoxin assays. More particularly, this invention relates to a homogeneous assay that uses changes in fluorescence polarization
10 to detect the presence of deoxynivalenol in grains.

2. Description of Related Art

Deoxynivalenol (DON), which is also known as vomitoxin, is a mycotoxin produced in various grains, such as wheat, corn, barley, oats and rye, by *Fusarium*
15 *graminearum* and other *Fusarium* strains¹⁻⁴. Presence of DON in food causes food refusal, vomiting and growth depression in swine, gastrointestinal illness in humans, and embryotoxicity and immunotoxicity in laboratory animals¹⁻⁴. Because of its latent health risks, research is being carried out to explore analytical methods of detecting DON.

More generally, DON is a particularly troublesome mycotoxin within the group of
20 mycotoxins known as trichothecenes. As shown in Figure 1, trichothecenes have a common skeleton that can have different groups attached at R¹-R⁵. Figure 2 identifies the R¹-R⁵ groups in Figure 1 for DON and for some of the other more well-known trichothecenes.

Various methods for the quantitative analysis of DON and other trichothecenes in
25 grains are in use, such as thin-layer chromatography (TLC), gas chromatography (GC),

and high-pressure liquid chromatography (HPLC). The TLC method is an official method (first action) of AOAC International. Although relatively simple, the TLC method lacks sensitivity and is only semiquantitative. In addition, most of these chromatographic methods require extensive cleanup procedures after extraction and are not suitable for field testing^{4,5}.

Recently, enzyme-linked immunosorbent assay (ELISA) methods have been successfully applied to the screening of DON in grains². However, ELISA methods are undesirably labor intensive, in that they typically involve several washings, liquid transfers, and incubation times. Accordingly, there is a need for a simple, yet sensitive, method for the determination of DON and other trichothecenes in grains that is rapid and field portable.

SUMMARY OF THE INVENTION

In a first principal aspect, the present invention provides a homogeneous assay for the determination of deoxynivalenol (DON) in grains. In accordance with the method, DON is extracted from a grain sample. The extract is combined with a tracer and an antibody to provide a mixture. The tracer comprises DON conjugated to a fluorophore, and the tracer is able to bind to the antibody to produce a detectable change in fluorescence polarization. The fluorescence polarization of the mixture is then measured. The measured fluorescence polarization is compared with a characterized fluorescence polarization value that corresponds to a known DON concentration.

In a second principal aspect, the present invention provides a homogeneous assay for the determination of trichothecenes in grains. In accordance with the method,

trichothecene is extracted from a grain sample. The extract is combined with a tracer and an antibody to provide a mixture. The tracer comprises a predetermined trichothecene conjugated to a fluorophore, and the tracer is able to bind to the antibody to produce a detectable change in fluorescence polarization. The fluorescence polarization of the mixture is then measured. The measured fluorescence polarization is compared with a characterized fluorescence polarization value that corresponds to a known trichothecene concentration.

In a third principal aspect, the present invention provides an assay kit for the determination of DON in grains. The assay kit comprises an antibody and a tracer, each in an amount suitable for at least one assay, and suitable packaging. The tracer comprises DON conjugated to a fluorophore, and the tracer is able to bind to the antibody to produce a detectable change in fluorescence polarization.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 shows the basic chemical structure of trichothecenes, with R^1 , R^2 , R^3 , R^4 , and R^5 representing substitution positions on the basic trichothecene skeleton.

Figure 2 is a table identifying the R^1 - R^5 groups in Figure 1 for some well-known trichothecenes.

Figure 3 is a standard curve for a fluorescence polarization assay for DON, using the data of Table 1, in accordance with a preferred embodiment of the present invention.

Figure 4 is a standard curve for a fluorescence polarization assay for DON, using the data of Table 2, in accordance with a preferred embodiment of the present invention.

Figure 5 is a standard curve for a fluorescence polarization assay for DON, using the data of Table 3, in accordance with a preferred embodiment of the present invention.

Figure 6 is a graph comparing the DON concentration of spiked samples with the DON concentration calculated from the standard curve of Figure 5, in accordance with a preferred embodiment of the present invention.

Figure 7 is a standard curve for a fluorescence polarization assay for DON, using the data of Table 5, in accordance with a preferred embodiment of the present invention.

Figure 8 is a graph comparing the DON concentration of samples as measured using HPLC with the DON concentration as calculated from the standard curve of Figure 7, in accordance with a preferred embodiment of the present invention.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The present invention provides a homogeneous assay for the determination of DON and its metabolites in grains that is based on measurements of fluorescence polarization. The technique of fluorescence polarization has been successfully utilized in various assays involving proteins, enzymes, therapeutic drugs, drugs of abuse, DNA, hormones, peptides and antibodies⁶⁻⁸.

The principle behind the fluorescence polarization technique is as follows. Fluorescent probes having low molecular weight have low polarization values due to their fast rotation, whereas fluorescent probes with higher molecular weight have higher polarization due to their slower rotation. Thus the polarization value of a fluorophore increases upon binding to a larger molecule. Further information about the fluorescence polarization technique is provided in U.S. Patent Nos. 5,427,960 and 5,976,820 and in

Nasir, M. S. and Jolley, M. E., "Fluorescence Polarization: An analytical tool for Immunoassay and Drug Discovery," *Combinatorial Chemistry & High Throughput Screening*, **1999**, 2, 177-190, which references are incorporated herein by reference.

The present invention uses a tracer, comprising a fluorophore conjugated to DON,
5 that provides a specified polarization value and is able to bind to an antibody specific to DON to produce a detectable change in fluorescence polarization. This DON-fluorophore tracer competes with free DON from grain extracts for binding to a specific antibody (an inhibition assay), giving rise to a fluorescence polarization value that is inversely proportional to the quantity of DON in the grain.

10 In preferred embodiments, the assay is rapid, simple (in that it requires minimal training), and field portable, making it useful for the routine and quantitative determination of DON in grains. The preferred assay can also detect very small amounts of DON (<0.2 PPM).

15 **1. Materials and Methods**

All the chemicals and solvents were used as received unless otherwise noted. DON and fluoresceinamine (isomer II) were obtained from Sigma. Monoclonal antibodies specific to DON were provided by Dr. Chris Maragos (USDA, Peoria, IL).
20 Reaction mixtures were separated on preparative TLC using silica gel plates (Sigma). Fluorescence polarization was measured at room temperature using a single tube Sentry-FP fluorescence polarization instrument (Diachemix Corp.). Data was plotted using the program Graph Pad Prism, and unknown values were obtained from the graph.

2. DON Monoclonal Antibody Preparation

A number of different techniques for preparing DON specific antibodies have been reported by various groups^{1-2,9}. Since DON is a small molecule, it is typically conjugated with a protein to be made immunogenic.

5 The preferred method for preparing DON specific antibodies, which was the method used to prepare the antibodies used in these studies, is as follows. DON (5 mg) is reacted with 1,1'-carbonyldiimidazole (40 mg) in 800 µl acetone for 1 hour at room temperature. Water (20 µl) is slowly added followed by 5.8 mg of ovalbumin in 385 µl of 0.1 M sodium bicarbonate buffer (pH 8.5). The mixture is kept at 4° C for 24 hours and dialyzed extensively against 0.1 M phosphate-buffered saline (PBS). The DON-protein conjugate is diluted to 1mg/ml with 0.1 M PBS, distributed as 200 µl portions, and lyophilized. This lyophilized material is reconstituted with water immediately before using it to immunize of mice. The antibodies produced by these cell lines are then screened for DON specificity. Further details are provided in Chris M. Maragos and Susan P. McCormick, "Monoclonal Antibodies for the Mycotoxins Deoxynivalenol and 3-Acetyl-Deoxynivalenol," *Food and Agricultural Immunology*, **2000**, 12, 181-192, which is incorporated herein by reference.

3. Preparation of DON-F Tracer

20 100 µL of an acetone solution of DON (Sigma, 0.625 mg) was mixed with 2 mg of 1,1 carbonyldiimidazole (Sigma) and kept at room temperature. After two hours, this was mixed with 100 µl of a solution of fluoresceinamine isomer II (6-aminofluorescein, Sigma, 10 mg/ml in 0.1 molar sodium carbonate, pH ~ 9.5) and after thorough shaking, 25 the reaction was incubated overnight at room temperature. The resultant product was

separated by preparative TLC (silica, CHCl₃: CH₃OH: CH₃CO₂H, 90: 10: 1), and the product (R_f ~ 0.4) was collected, shaken with methanol, centrifuged and filtered to give pure DON-F tracer. This stock solution of DON-F tracer was stored at 2-8° C and diluted appropriately for use. Other fluorophores could also be used, depending on the antibody.

5 It was observed that 10 µl of this diluted tracer in 1 ml PBS gave an intensity equivalent to ~ 1 nM fluorescein. Furthermore, this tracer was found to give a polarization of 40-50 mP, which changed to ~250 mP upon adding the appropriately diluted DON antibody provided by Dr. Chris Maragos (USDA, Peoria, IL).

10 4. Protocol for DON Fluorescence Polarization Assay

 A preferred protocol for performing the DON assay is as follows. The grain sample being tested for DON is crushed. Then, 20 grams of the crushed grain is shaken in 100 ml of water for 10-20 minutes to extract the DON for analysis. Beneficially, this
15 extract may be used without any additional purification steps.

 In order to determine the amount of DON present in the grain samples, a standard curve is first obtained using standard DON solutions as follows. A series of standard DON solutions may be made by diluting an appropriate amount of stock aqueous DON solution (10 mg/ml) into 1 ml PBSA buffer that contains bovine gamma globulin (BGG)
20 in a concentration of 0.01%. A fixed amount of standard, such as 10 µl or 20 µl, is pipetted into 1 ml of diluted antibody solution in a 10x75 mm glass test tube, and the mixture is vortexed thoroughly. The diluted antibody solution is preferably about 1/20,000 to 1/40,000 in PBSA with BGG. This mixture is used to perform a blank reading in the instrument. The readings should be repeated until they are stable

(normally two readings). 10 µl of DON-F tracer solution, prepared as described above, is then added to the glass test tube containing the antibody solution and standard, and the test tube is vortexed thoroughly. The test tube is placed back in the instrument, and mP values are recorded until they stabilize. The other standard solutions are read in the same way, and a standard curve is constructed using the stabilized mP values.

The samples, prepared as described above, are measured in a similar manner. Specifically, a fixed amount of extracted sample, typically 10 µl or 20 µl, depending on what was used for the standard curve, is pipetted into 1 ml of diluted antibody solution in a 10x75 mm glass test tube. The mixture is vortexed thoroughly and used to perform a blank reading in the instrument. 10 µl of tracer is added to the glass test tube containing the antibody solution and sample, and the test tube is vortexed thoroughly. The test tube is placed back in the instrument, and mP values are recorded. The DON concentration in the extracted sample is then calculated using the standard curve.

5. Standard Curve For Fluorescence Polarization Assay of DON

Standard curves were obtained for the Sentry-FP fluorescence polarization instrument using the protocol described above for DON standards in various concentrations. Table 1 lists the fluorescence polarization readings in mP units for DON standard solutions, using 10 µl of standard solution and “Antibody #1” from Dr. Chris Maragos (USDA, Peoria, IL). Figure 3 shows the standard curve, relating the fluorescence polarization signal in mP to DON concentration in ppm, that was obtained from this data.

TABLE 1

DON standard concentration (ppm)	mP
0.00	171
0.10	143
0.25	130
0.50	112
1.00	87
2.00	65
3.00	52
4.00	46
5.00	43

Table 2 lists the fluorescence polarization readings in mP units for DON standard solutions, using 20 μ l of standard solution and “Antibody #4” from Dr. Chris Maragos (USDA, Peoria, IL). Figure 4 shows the standard curve, relating the fluorescence polarization signal in mP to DON concentration in ppm, that was obtained from this data.

TABLE 2

DON standard concentration (ppm)	mP
0.00	247
0.10	241
0.25	215
0.50	183
1.00	145
2.00	98
4.00	76

6. Results of Fluorescence Polarization Assay For DON Spiked Wheat Samples

DON free wheat was supplied by Dr. Chris Maragos (USDA, Peoria, IL). 20 g wheat samples were supplemented with varying concentrations of DON (Sigma, 1 mg/ml stock solution) and extracted with 100 ml water by shaking for 10-20 minutes. 1 ml of

the supernatant was centrifuged in each case and analyzed. If not analyzed immediately, samples were stored in a refrigerator. 20 µl of sample was mixed with 1 ml of a 1/40,000 solution of “Antibody #1” from Dr. Chris Maragos (USDA, Peoria, IL) in PBSA-BGG buffer and blanked in the Sentry-FP instrument. 10 µl of properly diluted tracer was then added and the fluorescence polarization measured. Each sample was run in duplicate. A standard graph was plotted (Figure 5), using the data of Table 3. “Antibody #4,” however, was found not to yield reliable results with real samples.

TABLE 3

DON standard concentration (ppm)	mP (run 1)	mP (run 2)	mP (run 3)
0	224	228	223.45
0.5	203.08	192.33	196
1.25	168.89	171.49	169.02
2.5	143.2	145	146.42
5	111.12	110.07	112.01
10	82.68	84.92	86.52

The DON concentration (in ppm) of each sample was calculated from the standard graph of Figure 5, using the average mP value of the duplicate runs of that sample. Table 4 shows these results. Figure 6 is a plot of the DON concentration (in ppm) of the samples as they were spiked versus the DON concentration (in ppm) calculated from the standard curve of Figure 5. A good correlation (an r^2 value of 0.994) was observed between these theoretical and calculated DON concentrations.

TABLE 4

DON concentration of spiked sample (ppm)	mP	Calculated DON concentration (ppm)	% Recovery of DON
3.5	134.03	2.95	84.28
0.5	200.52	0.49	98
1.5	169.00	1.42	94.66
6.5	100.64	6.00	92.3
10	178.29	11.1	111
20	151.03	21	105
0.2	209.85	0.26	130
1.2	177.30	1.14	95
0.8	188.01	0.82	102
2.5	147.17	2.28	91.2
8	186.31	8.7	108

7. Comparison of Fluorescence Polarization Assay For DON With HPLC

A number of different wheat samples with varying amounts of DON, provided by Dr. Chris Maragos (USDA, Peoria, IL), were analyzed by the fluorescence polarization assay procedure described above using Antibody #1. The samples were also analyzed by HPLC using standard methods. Table 5 shows the data for the standard graph (Figure 7) used in this study.

TABLE 5

DON standard concentration (ppm)	mP (run 1)	mP (run 2)	mP (run 3)	mp (run 4)
0	235	235	234	232
0.2	225	228	222	222
0.5	214	217	215	214
1.25	201	201	203	203
2.5	190	190	191	191
5	161	161	164	163
10	134	135	133	133
20	107	107	108	109

Table 6 shows the DON concentration (in ppm) of each sample calculated from the standard graph of Figure 7.

TABLE 6

Sample Number	mP	Calculated DON Concentration (ppm)
1	133.5	9.46
2 (1x10 dilution)	196.75	1.93
3	182.25	3.06
4	139.5	8.26
5	144	7.48
6	125	11.64
7	123	12.27
8	136	8.93
9	110.5	18.73
10	153.25	6.12
11	133.5	9.46
12	156.25	5.73
13	228.5	0.04
14	234	0.00
15	121.5	12.79
16	126.75	11.13
17	129.75	10.33
18	146.25	7.12
19	166.5	4.55
20	173	3.89
21	190	2.43
22	159.75	5.30
23	168.5	4.34
24	237.75	0.00
25	170.75	4.11
26	199	1.77
27	213.5	0.82
28	217.5	0.58
29	212	0.92
30	155	5.89
31	213.25	0.84
32	161.75	5.07
33	228	0.05
34	184.25	2.89

Figure 8 is a plot of the calculated DON concentrations (in ppm) from the fluorescence polarization assay, as shown in Table 6, versus the DON concentration (in ppm) of the sample as measured by HPLC. A best fit line was calculated from this data. Its slope was 1.2112 ± 0.0391 , and the r^2 value was 0.9676, indicating that the results from the fluorescence polarization assay were in good agreement with the results from HPLC.

8. Other Trichothecenes

It has been found that Antibody #1, when used in a fluorescence polarization assay with the DON-F tracer as described above, has some cross-reactivity with other trichothecenes. Specifically, Antibody #1 was found to have a 358% cross-reactivity with 15-acetyl-deoxynivalenol and a 9% cross-reactivity with HT-2 toxin. The other trichothecenes listed in Figure 2 were found to have less than a 4% cross-reactivity.

The cross-reactivity percentage for each trichothecene was calculated by dividing the IC_{50} value for DON by the IC_{50} value of the trichothecene and multiplying by 100%. The IC_{50} value of a trichothecene is the concentration of the trichothecene required to give a fluorescence polarization response of 50%, where 100% corresponds to the response of the assay without the trichothecene and 0% represents the response of the assay without antibody.

Because of this cross-reactivity, the fluorescence polarization assay of the present invention may be used for the determination of trichothecenes other than DON. Moreover, because DON is closely related to other trichothecenes, fluorescence polarization assays may be developed specifically for these other trichothecenes.

9. Assay Kit

The materials used to perform the assay of the present invention are preferably made available in kit form. The kit preferably includes tracer and antibody in an amount suitable for at least one assay, along with suitable packaging and instructions for use. The tracer and antibody may be provided in solution, as a liquid dispersion, or as a substantially dry powder (e.g., in lyophilized form).

The suitable packaging can be any solid matrix or material, such as glass, plastic, paper, foil, and the like, capable of separately holding within fixed limits the buffer, tracer, and antibody. For example the tracer and monoclonal antibody may be provided as solutions in separate labeled bottles or vials made of glass or plastic.

The antibody is specific for DON and is preferably a monoclonal antibody. The preferred monoclonal antibody may be prepared as described herein and as known in the art.

The tracer comprises a fluorophore, such as 6-aminofluorescein, conjugated to DON. Other fluorophores may be used, provided the resulting tracer is able to bind with the monoclonal antibodies to produce a detectable change in fluorescence polarization.

10. References

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